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# Comparison of First-Trimester Screening Biomarkers in Spontaneous (Natural) and In Vitro Fertilization Pregnancies



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**Original Article** 

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#### Abstract

**Objectives:** The present study was designed to evaluate and compare the screening markers of spontaneous and in vitro fertilization (IVF) pregnancies in individuals who underwent combined screening in the first trimester of pregnancy.

Materials and Methods: In this retrospective study, we examined the difference between natural and assisted-reproductive technology (ART) pregnancies in 2252 mothers from January 2011 to October 2019. In both groups, the first trimester screening parameters including NT, free beta-human chorionic gonadotropin ( $\beta$ -hCG), and PAPP-A were measured.

**Results:** According to the results of the present study, BMI was higher in the group that became pregnant by IVF compared to the normal pregnancy group (P<0.001). Additionally, in patients using IVF, the history of maternal disease was less frequent compared to the other group (P=0.003). Most cases of twin fetuses, dichorionic twin pregnancy, and fetal abnormality were seen in the IVF group. Although the  $\beta$ -hCG and free PAPP-A levels were high in the IVF group, no significant difference was observed. The group with spontaneous pregnancy had a higher penetration rate of nuchal translucency <95th percentile compared to the IVF group (P<0.001). The results of comparing the first trimester test showed that in the IVF group, 81.3% of patients were at low risk and 6.6% were at high risk, while in the normal pregnancy group, 78.9% of patients were at low risk and 7.0% of them were at low risk, indicating that the difference was not significant.

**Conclusions:** Overall, although the risk of an euploidy in IVF pregnancies was not higher compared to normal pregnancies, in IVF pregnancies, NI is associated with a high risk of fetal abnormalities. Therefore, NT sonography can help to diagnose fetal anomalies especially an euploidy.

Keywords: IVF, first-trimester screening, free β-HCG, PAPP-A, NT

### Introduction

The use of ultrasound markers and maternal serum samples during pregnancy are two important methods for screening along with other maternity care worldwide. Maternal age, nuchal translucency (NT), free beta-human chorionic gonadotropin ( $\beta$ -hCG) level, and maternal serum level of PAPP-A in the first trimester of pregnancy are the main tools in diagnosing Down syndrome and other trisomies (1). Based on the results of studies, the rate of false-positive results of this method are reported to be about 5% and approximately 90% of trisomy 21 cases can be detected by this method.

The assisted-reproductive technologies (ART) annually lead to a large number of pregnancies. The age of the women who undergo assisted reproductive treatment for their pregnancies is higher than the average age. Therefore, the risk of Down syndrome and other aneuploidies is higher in them (2, 3). Based on various research works, there is an association between pregnancies achieved by ART and alteration in the first trimester screening biomarkers, which influences the assessment of the risk of Down syndrome (4-6). The application of an efficient screening test with a low rate of false-positive results is needed in

these patients. It has been shown that the combination of such parameters as NT, mother's age,  $\beta$  -hCG, and PAPP-A in first-trimester screening helps identify almost 85%-90% of trisomy 21 cases and other aneuploidies, with a screen positive rate of 5%-6% (1,7). The reduced maternal serum level of PAPP-A and normal or higher  $\beta$ -hCG levels in vitro fertilization (IVF) pregnancies are the most consistent results in these research works, which was found by a new meta-analysis in this regard (2,6,8). However, findings of some other studies showed that there is not any difference between IVF and natural pregnancies in terms of these biomarkers (9,10). The inconsistency among the research findings is apparently due to some confounding factors, including adverse obstetric outcomes, maternal age, and non-homogeneous patient populations. Hence, the present research attempted to examine the probable impact of ART on first-trimester screening for aneuploidy through controlling fetal NT and PAPP-A levels and the maternal serum concentration of free  $\beta$ -HCG.

#### **Patients and Methods**

The population of this study included pregnant women in the first trimester of pregnancy who became pregnant

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#### Key Messages

- There was no difference in the risk of an uploidy between normal and IVF pregnancies.
- In IVF pregnancies, NT levels were associated with a high risk for fetal abnormalities.
- ► There was no difference in biochemical markers of aneuploidy between normal and IVF pregnancies.

spontaneously and those who used assisted reproductive techniques. Assisted reproductive techniques included ovulation induction (OI), intrauterine insemination (IUI), and IVF. The mentioned samples who referred to a sub-specialized center in Ahvaz from January 2011 to October 2019 were examined

All patients referred during this period were included in the study. Participants who were dissatisfied with the study and cases whose information was incomplete were excluded from the study. Totally, 2252 mothers were evaluated, including 1879 spontaneous and 373 IVF pregnancies.

Data were entered into a database containing demographic information, ultrasound findings, and biochemical test results which were examined and specific hazards were identified.

Ultrasound specialists certified by the Fetal Medical Foundation evaluated fetal NT in the first trimester of pregnancy using the abdominal ultrasound methods like the maximum vertical distance between the skin and subcutaneous tissues of the back of the neck in a sagittal section of the fetus in the neutral position. Screening for chromosomal abnormalities was done using three methods: maternal age, fetal NT, and concentrations of  $\beta$ -HCG and PAPP-A.

For the assessment of an uploidy (trisomy 21, trisomy 18, and trisomy 13), we used amniocentesis and chorionic villus sampling and subsequent karyotyping or cell-free DNA test.

### Statistical Analysis

Data analysis was performed using GraphPad Prism version 7. Descriptive statistics were used to describe the demographic characteristics, which included maternal age, body weight, gestational age, and BMI (mean  $\pm$  standard deviation). Mean, SD, and percentile were also used to describe the MoM (multiple of median) screening markers of the first trimester (NT,  $\beta$ -hCG, and PAPP-A). *T*-test and Mann-Whitney U test were used to express the monthly mean differences between spontaneous and IVF pregnancies. The relationship between variables in the first-trimester screening test was performed using bivariate Pearson correlation test and linear regression analysis. *P*<0.05 was considered statistically significant.

### Results

According to the results of the study, there was no

significant difference between the mean age of the IVF pregnancy group and the normal pregnancy group (P>0.05). The mean weight of patients in the IVF pregnancy group was lower than the other group but BMI was higher (P<0.001) (Table 1).

#### **Clinical Characteristics**

The frequency of cases with a history of maternal disease among IVF group was statistically less than those with normal pregnancy (P=0.003).

In contrast, the prevalence of twin fetuses, dichorionic twins, and fetal abnormalities was higher in the IVF group (Table 2).

# First Trimester Screening Biomarkers in Spontaneous and IVF Pregnancies

The mean  $\beta$ -hCG and free PAPP.A levels in the IVF pregnancy group were higher but not significantly. In the normal pregnancy group, the penetration rate of NT was less than 95th percentile compared to the IVF group (*P*<0.001) (Table 3).

Table 1. Demographic Characteristics of the Participants in Two Groups (Mean  $\pm$  SD)

Variables	Spontaneous Pregnancy	IVF Pregnancy	P Value
Number	1879	373	
Age	$30.13 \pm 4.86$	$30.17 \pm 5.53$	0.885
BMI	$26.17 \pm 4.43$	$27.18 \pm 4.28$	< 0.001
Gravidity	1.91 ± 1.07	$1.50\pm0.87$	< 0.001

BMI, Body mass index; IVF, In vitro fertilization.

Fable 2. Clinical Characteri	tics of the Participa	ants in the Two Groups
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Variables	Spontaneous Pregnancy	IVF Pregnancy	P Value
History of maternal disease			0.003
Yes	238 (22.1%)	15 (11.1%)	
No	837 (77.9%)	121 (89.9%)	
P value	< 0.0001	< 0.0001	
Fetus			< 0.001
Single	1862 (99.1%)	354 (94.9%)	
Twin	17 (0.9%)	19 (5.1%)	
P value	< 0.0001	< 0.0001	
Chorionicity			0.010
Monochorionic	9 (52.9%)	2 (10.5%)	
Dichorionic	8 (47.1%)	17 (89.5%)	
P value	0.157	0.001	
History of fetal trisomy <sup>a</sup>			0.101
Yes	25 (2.3%)	0 (0%)	
No	1059 (97.7%)	136 (100%)	
P value	< 0.0001	-	
Anomaly in sonography <sup>b</sup>			< 0.001
Yes	2 (0.1%)	15 (3.3%)	
No	537 (99.9%)	439 (96.7%)	
<i>P</i> value	< 0.0001	< 0.0001	

SD: Standard deviation, BMI: Body mass index, IVF: In vitro fertilization.

<sup>a</sup> Trisomy 21, trisomy 18, or trisomy 13.

<sup>b</sup> Any structural anomaly reported in the first-trimester ultrasound.

# Relationship Between First Trimester Screening Biomarkers and Clinical Characteristics

In screen-positive cases, the mean Free  $\beta$ -hCG level was significantly higher compared to screen-negative cases (*P*=0.016). The mean PAPP-A level was higher in single pregnancy than in twin pregnancy (*P*=0.071). However, the mean PAPP-A level in cases with a positive history of fetal trisomy was lower than in cases with a negative history of fetal trisomy. Table 4 shows the association between NT and independent variables. NT less than 95th percentile was more likely among cases with a history of maternal disease (*P*=0.009) and single fetus (*P*<0.001) (Table 4).

#### Distribution of the Risk for Aneuploidy

In the IVF group, 81.3% of patients were at low risk and 6.6% were at high risk. Additionally, in the group with normal pregnancy, 78.9% were at low risk and 7% were at high risk, which was not a significant difference (P = 0.609) (Table 5).

#### Demographic Characteristics According to Risk Factor

The distribution of independent variables and covariates in three risk categories is presented in Table 6. It was shown that mean age, mean gravidity, and mean BMI were statistically lower among low-risk groups of cases (P < 0.001). Moreover, the rate of anomaly was statistically high in the high-risk group (P < 0.001) (Table 6).

 Table 3. First Trimester Screening Biomarker in Spontaneous and IVF

 Pregnancies

Variables	None IVF	IVF	P Value
Free β-hCG	$1.19\pm0.89$	$1.21 \pm 0.69$	0.822
PAPP-A	$1.18\pm0.68$	$1.31 \pm 0.93$	0.060
NT			
(>95 <sup>th</sup> centile)	102 (5.4%)	61 (16.4%)	-0.001
(<95 <sup>th</sup> centile)	1777 (94.6%)	312 (83.6%)	<0.001

SD: Standard deviation, BMI: Body mass index, IVF: In vitro fertilization, NT: nuchal translucency.

Table 4. The Association	of Free β-hCG	and PAPP.A with	Independent Variables
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# First Trimester Screening Biomarkers according to Risk Factor

The rate of fetal malformations was higher in the high-risk group than in the low-risk and moderate groups. There was no difference between the three groups in terms of maternal disease and history of fetal trisomy in previous pregnancies (Table 7).

## Discussion

The findings of the present research indicated a significantly higher BMI in the IVF pregnancy group compared to the natural pregnancy group (P < 0.001). As found by Rittenberg et al, the live birth rate using ART was lower in women having a high BMI than women who had a normal BMI (11). Various other studies, which evaluated the effect of BMI on ART results, supported this finding (12-14). Mortality, genital diseases, infertility, and amenorrhea are significantly related to obesity. Moreover, the probability of experiencing menopausal disorders in obese women is three times higher compared to women with normal weight. The impact of obesity on the quality of eggs, embryo growth, implantation rate, and number of mature eggs has been reported in various studies. The success rate of assisted reproductive techniques is lower in obese women compared to women with normal BMI, as shown by different research works. It can be justified by the relationship between obesity and lower estradiol levels and the lower number of fertilized oocytes (15-17).

In this study, the birth rate of twins was significantly higher in the IVF pregnancy group compared to the natural pregnancy group (P=0.003). A total of 582 normal pregnancy cases and 2414 ART pregnancy cases were studied by Levi Setti et al, and it was found that twin and triplet birth rates were significantly higher in the ART pregnancy group compared to the normal pregnancy group (18). Approximately 77% of triplet and higherorder births (25% IVF birth and 52% non-IVF birth) and 40% of twin births (19% IVF birth and 21% non-IVF birth) are estimated to be accounted for assisted birth (19,

		PAPP.A	Free β-hCG	NT (>95th Percentile)	NT (<95th Percentile)
	No	$1.21 \pm 0.72$	$1.19 \pm 0.86$	86 (9%)	872 (91%)
History of maternal disease	Yes	$1.16 \pm 0.70$	$1.21 \pm 0.90$	10 (4%)	243 (96%)
	P value	0.269	0.836	0.00	9
	Single	$1.19 \pm 0.72$	$1.19 \pm 0.88$	153 (6.9%)	2063 (93.1%)
Fetus	Twin	$1.41 \pm 0.60$	$1.20 \pm 0.70$	10 (27.8%)	26 (72.2%)
	P value	0.071	0.962	<0.00	01
	No	$1.22 \pm 0.68$	$1.26 \pm 0.97$	93 (7.8%)	1102 (92.2%)
History of fetal trisomy <sup>a</sup>	Yes	$0.87 \pm 0.58$	$1.02 \pm 0.72$	3 (12%)	22 (88%)
	P value	0.001	0.130	0.43	8
Anomaly in sonography <sup>b</sup>	No	$1.31 \pm 0.7$	$0.86 \pm 0.39$	26 (70.27 %)	519 (98.85%)
	Yes	$1.45 \pm 0.56$	$1.35 \pm 0.76$	11 (29.73%)	6 (1.14%)
	P value	0.582	0.016	<0.00	01

<sup>a</sup> Trisomy 21, trisomy 18, or trisomy 13.

<sup>b</sup> Any structural anomaly reported in the first-trimester ultrasound.

Category	Spontaneous Pregnancy	IVF Pregnancy	P Value
Low	840 (78.9%)	74 (81.3%)	
Intermediate	150 (14.1%)	11 (12.1%)	0.000
High	74 (7.0%)	6 (6.6%)	0.609
P value	< 0.0001	< 0.0001	

Table 6. The Association of Risk and Independent Variables

Parameter	Low	Intermediate	High
Age	$29.79 \pm 4.32$	$33.66 \pm 5.20^{a}$	$33.61 \pm 4.45^{a}$
BMI	$25.86 \pm 4.28$	$27.36 \pm 4.25^{a}$	$27.55 \pm 4.53^{a}$
Gravidity	$1.97 \pm 1.14$	$2.49 \pm 1.41^{\text{a}}$	$2.43 \pm 1.24^{a}$

<sup>a</sup> Significant difference with Low risk group.

20). According to the findings of recent studies, maternal age, young oocyte age, number of high-quality embryos transferred, treatment period, obesity, and higher-quality of embryos transferred are associated with a higher possibility of twin birth after IVF (21-24).

In our study, the rates of congenital abnormalities and dichorionic twins were higher in the IVF group compared to the natural pregnancy group. Dizygotic (DZ) twins are dichorionic (DC), while monozygotic (MZ) twins might be DC or monochorionic (MC). Regardless of some deficiencies resulting from intrauterine crowding such as deformities in foot, skull asymmetry, and hip dislocation, the abnormality rate in singletons is similar to that in DZ twins, while the abnormality rates among MZ twins are higher. In addition, mortality rates among MZ twins are higher. These higher risks have been indicated to be limited to MC MZ twins, while DC MZ and DZ twins show similar results (25-27).

Due to double-embryo transfer, most of ART twin pairs are DZ. However, MZ twinning from embryo splitting is concerned in ART twin conceptions too. More recent studies have reported higher MZ twinning rates (1%-5%) among ART conceptions compared to the rates seen in normal conceptions (0.4%) (28-30). The age of infertile couples, the main cause of infertility, the medicines taken for induced ovulation or for maintaining the pregnancy in the first months, as well as the procedure-related factors, such as the delayed fertilization, the embryo freezing, and defrosting, and the polyspermic fertilization potential are among the factors that increase the risk of birth abnormalities (31).

It is theoretically logical to assume that IVF directly influences the first-trimester screening, and it is especially affected by the presence of multiple corpora lutea and hormonal treatments. Hence, evident serum marker alterations can be expected during the first trimester. These changes are caused by this treatment (32). However, IVF and natural conceptions showed no significant differences in terms of the PAPP-A levels in the first trimester in the present study. According to previous studies, ART protocols did not influence PAPP-A serum levels in the first trimester (10,32). Following IVF, some impacts on serum markers were found in the first trimester, i.e., reduced PAPP-A level (9,33,34). Decreased PAPP-A levels indicate a decrease in insulin growth factor in Down syndrome pregnancy (35,36).

Given the involvement of IGFs in trophoblast invasion, it is possible to associate the alterations in the bioavailability of these hormones with the higher prevalence of pregnancy complications related to Downs' syndrome conceptions. The reduced maternal serum level of PAPP-A in Down's syndrome conceptions is not specific since low PAPP-A levels are also observed in pre-eclampsia and intrauterine growth retardation pregnancies (37,38).

The  $\beta$ -hCG is known as one of the biochemical markers in the first-trimester screening. Despite the increase or reduction of  $\beta$ -hCG MoM levels in ART conceptions in comparison with control groups reported by some research works (5,6,39,40), most studies, as well as the present study, did not show any difference between ART and natural conceptions in terms of  $\beta$ -hCG levels (2,4,9,41). The inconsistency among the findings is due to small sample sizes, the heterogeneous research populations, and different levels of  $\beta$ -hCG at different pregnancy weeks (4,41). For a long time, free  $\beta$ -hCG has been utilized as a chemical marker in early pregnancy,

Table 7. The Association of Risk and Independent Variables

Parameter		Low	Intermediate	High	P Value
	No	721 (80.2%)	120 (13.3%)	58 (6.5%)	
History of maternal	Yes	191 (75.4%)	41 (16.2%)	21 (8.3%)	0.248
disease	P value	<0.0001	< 0.0001	< 0.0001	
History of fetal trisomy <sup>a</sup>	No	900 (79.4%)	157 (13.8%)	77 (6.8%)	
	Yes	14 (66.7%)	4 (19%)	3 (14.3%)	0.288
	P value	<0.0001	< 0.0001	< 0.0001	
Anomaly in sonography <sup>b</sup>	No	444 (98%)	63 (92.6%)	29 (90%)	
	Yes	9 (1.98%)	5 (7.35%)	3 (9.3%)	< 0.001
	P value	<0.0001	< 0.0001	0.004	

<sup>a</sup> Trisomy 21, trisomy 18, or trisomy 13.

<sup>b</sup> Any structural anomaly reported in the first-trimester ultrasound.

and the free  $\beta$ -subunit has been identified as having a higher predictive value for Down's syndrome. However, not all studies have confirmed this claim (42,43). Free  $\beta$ -hCG, a vital marker for pregnancy maintenance, is generated by trophoblast. It is assumed that the hCG level in early pregnancy is representative of the mass of syncytial trophoblast (44). For the purpose of screening Down's syndrome, measurement of free  $\beta$ -hCG is done at 8–14 weeks of pregnancy. The high free  $\beta$ -hCG levels in the first trimester are related to Down's syndrome, and they are associated with poor obstetric outcomes in the second trimester (38). The free  $\beta$ -hCG level is lower in the first trimester in IVF pregnancies compared to natural conceptions. However, the level of free  $\beta$ -hCG in the second trimester is higher in IVF conceptions (44,45).

The single ultrasound marker in the first trimester combined screening procedure is NT. As indicated by the current research, NT thickness is different in ART conception and natural conception. These findings are in contrast to the findings of other research concerning the thickness of NT in ART pregnancies. Previous studies did not report any significant difference in the NT in ICSI, IVF conceptions, or ovulation induction (10,32). According to a study conducted in Italy, which consisted of 32 IVF and 42 ICSI conceptions, there was not any difference in NT (9). In a cohort study conducted in Israel, with a smaller sample size, a non-significant rise was found in NT measurements (46,47). Nevertheless, the finding of the present study is consistent with a previous study indicating increased NT in IUI conceptions compared to natural conceptions (48). The basic pathophysiology for higher NT thickness is not still clear. The mechanisms reported for this include changed the composition of the extracellular matrix of the nuchal skin, cardiac failure, and delayed or abnormal growth of the lymphatic system (49-51). There is an association between increased NT and syndromal, chromosomal, and structural anomalies (52, 53).

## Conclusions

To sum up, there was no significant difference in  $\beta$ -hCG and PAPP.A levels between the two groups with IVF and spontaneous pregnancies, but there was a significant difference in NT measurements. On the other hand, there are significant differences between IVF pregnancies and spontaneous pregnancies. It can be concluded that the risk of aneuploidy in IVF pregnancies is not higher compared to normal pregnancies, However, In IVF pregnancy, NT was associated with a high risk for fetal anomalies. Therefore, NT sonography can help to diagnose fetal anomalies, especially aneuploidy.

#### **Authors' Contribution**

Study concept and design: MB and MZ; analysis and interpretation of data: MB, MZ, and BA; drafting of the manuscript: BA; critical revision of the manuscript for important intellectual content: MB and MZ.

### Conflict of Interests

Authors have no conflict of interests.

#### **Ethical Issues**

This study was approved by the Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran (ethics code: IR.AJUMS. REC.1398.780).

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#### References

- Spencer K, Spencer CE, Power M, Dawson C, Nicolaides KH. Screening for chromosomal abnormalities in the first trimester using ultrasound and maternal serum biochemistry in a onestop clinic: a review of three years prospective experience. BJOG. 2003;110(3):281-286.
- Gjerris AC, Loft A, Pinborg A, Christiansen M, Tabor A. First-trimester screening markers are altered in pregnancies conceived after IVF/ICSI. Ultrasound Obstet Gynecol. 2009;33(1):8-17. doi:10.1002/uog.6254
- Anckaert E, Schiettecatte J, Sleurs E, Devroey P, Smitz J. First trimester screening for Down's syndrome after assisted reproductive technology: non-male factor infertility is associated with elevated free beta-human chorionic gonadotropin levels at 10-14 weeks of gestation. Fertil Steril. 2008;90(4):1206-1210. doi:10.1016/j.fertnstert.2007.08.050
- 4. Amor DJ, Xu JX, Halliday JL, et al. Pregnancies conceived using assisted reproductive technologies (ART) have low levels of pregnancy-associated plasma protein-A (PAPP-A) leading to a high rate of false-positive results in first trimester screening for Down syndrome. Hum Reprod. 2009;24(6):1330-1338. doi:10.1093/humrep/dep046
- Engels MA, Kooij M, Schats R, Twisk JW, Blankenstein MA, van Vugt JM. First-trimester serum marker distribution in singleton pregnancies conceived with assisted reproduction. Prenat Diagn. 2010;30(4):372-377. doi:10.1002/pd.2495
- Bender F, Hecken J, Reinsberg J, et al. Altered first-trimester screening markers after IVF/ICSI: no relationship with smallfor-gestational-age and number of embryos transferred. Reprod Biomed Online. 2010;20(4):516-522. doi:10.1016/j. rbmo.2009.12.025
- Krantz DA, Hallahan TW, Orlandi F, Buchanan P, Larsen JW Jr, Macri JN. First-trimester Down syndrome screening using dried blood biochemistry and nuchal translucency. Obstet Gynecol. 2000;96(2):207-213. doi:10.1016/s0029-7844(00)00881-4
- Cavoretto P, Giorgione V, Cipriani S, et al. Nuchal translucency measurement, free β-hCG and PAPP-A concentrations in IVF/ ICSI pregnancies: systematic review and meta-analysis. Prenat Diagn. 2017;37(6):540-555. doi:10.1002/pd.5052
- Orlandi F, Rossi C, Allegra A, et al. First trimester screening with free beta-hCG, PAPP-A and nuchal translucency in pregnancies conceived with assisted reproduction. Prenat Diagn. 2002;22(8):718-721. doi:10.1002/pd.390
- Wøjdemann KR, Larsen SO, Shalmi A, Sundberg K, Christiansen M, Tabor A. First trimester screening for Down syndrome and assisted reproduction: no basis for concern. Prenat Diagn. 2001;21(7):563-565. doi:10.1002/pd.124
- Rittenberg V, Seshadri S, Sunkara SK, Sobaleva S, Oteng-Ntim E, El-Toukhy T. Effect of body mass index on IVF treatment outcome: an updated systematic review and meta-analysis. Reprod Biomed Online. 2011;23(4):421-439. doi:10.1016/j. rbmo.2011.06.018
- 12. Zander-Fox DL, Henshaw R, Hamilton H, Lane M. Does obesity really matter? the impact of BMI on embryo quality

and pregnancy outcomes after IVF in women aged  $\leq 38$  years. Aust N Z J Obstet Gynaecol. 2012;52(3):270-276. doi:10.1111/j.1479-828X.2012.01453.x

- 13. Provost MP, Acharya KS, Acharya CR, et al. Pregnancy outcomes decline with increasing body mass index: analysis of 239,127 fresh autologous in vitro fertilization cycles from the 2008-2010 Society for Assisted Reproductive Technology registry. Fertil Steril. 2016;105(3):663-669. doi:10.1016/j. fertnstert.2015.11.008
- 14. Jungheim ES, Schon SB, Schulte MB, DeUgarte DA, Fowler SA, Tuuli MG. IVF outcomes in obese donor oocyte recipients: a systematic review and meta-analysis. Hum Reprod. 2013;28(10):2720-2727. doi:10.1093/humrep/det292
- Maheshwari A, Stofberg L, Bhattacharya S. Effect of overweight and obesity on assisted reproductive technologya systematic review. Hum Reprod Update. 2007;13(5):433-444. doi:10.1093/humupd/dmm017
- Fedorcsák P, Dale PO, Storeng R, et al. Impact of overweight and underweight on assisted reproduction treatment. Hum Reprod. 2004;19(11):2523-2528. doi:10.1093/humrep/ deh485
- 17. Bellver J, Ayllón Y, Ferrando M, et al. Female obesity impairs in vitro fertilization outcome without affecting embryo quality. Fertil Steril. 2010;93(2):447-454. doi:10.1016/j. fertnstert.2008.12.032
- Levi Setti PE, Moioli M, Smeraldi A, et al. Obstetric outcome and incidence of congenital anomalies in 2351 IVF/ ICSI babies. J Assist Reprod Genet. 2016;33(6):711-717. doi:10.1007/s10815-016-0714-4
- Sunderam S, Kissin DM, Crawford SB, et al. Assisted reproductive technology surveillance—United States, 2012. MMWR Surveill Summ. 2015;64(6):1-29.
- 20. Kulkarni AD, Jamieson DJ, Jones HW Jr, et al. Fertility treatments and multiple births in the United States. N Engl J Med. 2013;369(23):2218-2225. doi:10.1056/NEJMoa1301467
- Kim MS, Kim JH, Jee BC, Suh CS, Kim SH. Factors affecting occurrence of twin pregnancy after double embryo transfer on day 3. J Obstet Gynaecol Res. 2015;41(8):1223-1228. doi:10.1111/jog.12687
- 22. Niu ZH, Feng Y, Zhang AJ, Zhang HQ, Sun YJ, Lu XW. [Factors related to occurrence of twin pregnancy after double-embryo transfer in vitro fertilization cycles]. Zhonghua Fu Chan Ke Za Zhi. 2009;44(6):413-417.
- 23. Xu WH, Tong XM, Zhu HY, Lin XN, Jiang LY, Zhang SY. [Risk factors associated with twin pregnancy in double embryo transfer]. Zhonghua Yi Xue Za Zhi. 2011;91(37):2615-2618.
- Kaser DJ, Missmer SA, Correia KF, Ceyhan ST, Hornstein MD, Racowsky C. Predictors of twin live birth following cryopreserved double embryo transfer on day 3. J Assist Reprod Genet. 2013;30(8):1023-1030. doi:10.1007/s10815-013-0039-5
- Dubé J, Dodds L, Armson BA. Does chorionicity or zygosity predict adverse perinatal outcomes in twins? Am J Obstet Gynecol. 2002;186(3):579-583. doi:10.1067/ mob.2002.121721
- Minakami H, Sayama M, Honma Y, et al. Lower risks of adverse outcome in twins conceived by artificial reproductive techniques compared with spontaneously conceived twins. Hum Reprod. 1998;13(7):2005-2008. doi:10.1093/ humrep/13.7.2005
- 27. Loos R, Derom C, Vlietinck R, Derom R. The East Flanders Prospective Twin Survey (Belgium): a population-based register. Twin Res. 1998;1(4):167-175. doi:10.1375/136905298320566131
- Blickstein I, Verhoeven HC, Keith LG. Zygotic splitting after assisted reproduction. N Engl J Med. 1999;340(9):738-739.

doi:10.1056/nejm199903043400916

- Schachter M, Raziel A, Friedler S, Strassburger D, Bern O, Ron-El R. Monozygotic twinning after assisted reproductive techniques: a phenomenon independent of micromanipulation. Hum Reprod. 2001;16(6):1264-1269. doi:10.1093/humrep/16.6.1264
- Alikani M, Cekleniak NA, Walters E, Cohen J. Monozygotic twinning following assisted conception: an analysis of 81 consecutive cases. Hum Reprod. 2003;18(9):1937-1943. doi:10.1093/humrep/deg369
- 31. Tandulwadkar S, Lodha P, Kharb V. Congenital malformations and assisted reproductive technique: where is assisted reproductive technique taking us? J Hum Reprod Sci. 2012;5(3):244-247. doi:10.4103/0974-1208.106334
- Liao AW, Heath V, Kametas N, Spencer K, Nicolaides KH. First-trimester screening for trisomy 21 in singleton pregnancies achieved by assisted reproduction. Hum Reprod. 2001;16(7):1501-1504. doi:10.1093/humrep/16.7.1501
- Bersinger NA, Ødegård RA. Second- and third-trimester serum levels of placental proteins in preeclampsia and smallfor-gestational age pregnancies. Acta Obstet Gynecol Scand. 2004;83(1):37-45.
- 34. Hui PW, Lam YH, Tang MH, Ng EH, Yeung WS, Ho PC. Maternal serum pregnancy-associated plasma protein-A and free beta-human chorionic gonadotrophin in pregnancies conceived with fresh and frozen-thawed embryos from in vitro fertilization and intracytoplasmic sperm injection. Prenat Diagn. 2005;25(5):390-393. doi:10.1002/pd.1169
- 35. Giudice LC, Conover CA, Bale L, et al. Identification and regulation of the IGFBP-4 protease and its physiological inhibitor in human trophoblasts and endometrial stroma: evidence for paracrine regulation of IGF-II bioavailability in the placental bed during human implantation. J Clin Endocrinol Metab. 2002;87(5):2359-2366. doi:10.1210/jcem.87.5.8448
- 36. Santolaya-Forgas J, De Leon JA, Cullen Hopkins R, Castracane VD, Kauffman RP, Sifuentes GA. Low pregnancy-associated plasma protein-a at 10(+1) to 14(+6) weeks of gestation and a possible mechanism leading to miscarriage. Fetal Diagn Ther. 2004;19(5):456-461. doi:10.1159/000079000
- 37. Pihl K, Larsen T, Krebs L, Christiansen M. First trimester maternal serum PAPP-A, beta-hCG and ADAM12 in prediction of small-for-gestational-age fetuses. Prenat Diagn. 2008;28(12):1131-1135. doi:10.1002/pd.2141
- Dugoff L, Hobbins JC, Malone FD, et al. Quad screen as a predictor of adverse pregnancy outcome. Obstet Gynecol. 2005;106(2):260-267.
- Bersinger NA, Wunder D, Vanderlick F, et al. Maternal serum levels of placental proteins after in vitro fertilisation and their implications for prenatal screening. Prenat Diagn. 2004;24(6):471-477. doi:10.1002/pd.910
- 40. Lambert-Messerlian G, Dugoff L, Vidaver J, et al. Firstand second-trimester Down syndrome screening markers in pregnancies achieved through assisted reproductive technologies (ART): a FASTER trial study. Prenat Diagn. 2006;26(8):672-678. doi:10.1002/pd.1469
- Engels MA, Pajkrt E, Groot DT, Schats R, Twisk JW, van Vugt JM. Validation of correction factors for serum markers for firsttrimester Down syndrome screening in singleton pregnancies conceived with assisted reproduction. Fetal Diagn Ther. 2013;34(4):217-224. doi:10.1159/000355527
- 42. Shiefa S, Amargandhi M, Bhupendra J, Moulali S, Kristine T. First trimester maternal serum screening using biochemical markers PAPP-A and free  $\beta$ -hCG for Down syndrome, Patau syndrome and Edward syndrome. Indian J Clin Biochem. 2013;28(1):3-12. doi:10.1007/s12291-012-0269-9
- 43. Perni SC, Predanic M, Friedman A, Jean-Pierre C, Kalish RB,

Chasen ST. First-trimester screening with free  $\beta$ -hCG, PAPP-A, and nuchal translucency in pregnancies conceived by in vitro fertilization. Am J Obstet Gynecol. 2004;191(6):S44. doi:10.1016/j.ajog.2004.10.029

- 44. Almog B, Al-Shalaty J, Sheizaf B, et al. Difference between serum beta-human chorionic gonadotropin levels in pregnancies after in vitro maturation and in vitro fertilization treatments. Fertil Steril. 2011;95(1):85-88. doi:10.1016/j. fertnstert.2010.05.041
- 45. Bar-Hava I, Yitzhak M, Krissi H, et al. Triple-test screening in in vitro fertilization pregnancies. J Assist Reprod Genet. 2001;18(4):226-229. doi:10.1023/a:1009455912670
- 46. Maymon R, Shulman A. Serial first- and second-trimester Down's syndrome screening tests among IVF-versus naturallyconceived singletons. Hum Reprod. 2002;17(4):1081-1085. doi:10.1093/humrep/17.4.1081
- Braithwaite JM, Morris RW, Economides DL. Nuchal translucency measurements: frequency distribution and changes with gestation in a general population. Br J Obstet Gynaecol. 1996;103(12):1201-1204. doi:10.1111/j.1471-0528.1996.tb09629.x
- Lai TH, Chen SC, Tsai MS, Lee FK, Wei CF. First-trimester screening for Down syndrome in singleton pregnancies achieved by intrauterine insemination. J Assist Reprod Genet.

2003;20(8):327-331. doi:10.1023/a:1024813708875

- Böhlandt S, von Kaisenberg CS, Wewetzer K, Christ B, Nicolaides KH, Brand-Saberi B. Hyaluronan in the nuchal skin of chromosomally abnormal fetuses. Hum Reprod. 2000;15(5):1155-1158. doi:10.1093/humrep/15.5.1155
- Haak MC, Bartelings MM, Jackson DG, Webb S, van Vugt JM, Gittenberger-de Groot AC. Increased nuchal translucency is associated with jugular lymphatic distension. Hum Reprod. 2002;17(4):1086-1092. doi:10.1093/humrep/17.4.1086
- 51. Haak MC, van Vugt JM. Pathophysiology of increased nuchal translucency: a review of the literature. Hum Reprod Update. 2003;9(2):175-184. doi:10.1093/humupd/dmg008
- 52. Souka AP, Snijders RJ, Novakov A, Soares W, Nicolaides KH. Defects and syndromes in chromosomally normal fetuses with increased nuchal translucency thickness at 10-14 weeks of gestation. Ultrasound Obstet Gynecol. 1998;11(6):391-400. doi:10.1046/j.1469-0705.1998.11060391.x
- 53. Snijders RJ, Noble P, Sebire N, Souka A, Nicolaides KH. UK multicentre project on assessment of risk of trisomy 21 by maternal age and fetal nuchal-translucency thickness at 10-14 weeks of gestation. Fetal Medicine Foundation First Trimester Screening Group. Lancet. 1998;352(9125):343-346. doi:10.1016/s0140-6736(97)11280-6

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