Determination of Factor II Codons Genotype in Southeastern Iranian Patients With Hereditary Deficiency of Factor II

Hamed Soleimani Samarkhazan1, Shaban Alizadeh1*, Ziba Majidi2, Zahra Kashani Khatib3, Majid Naderi4

Abstract
Objectives: Congenital prothrombin (factor II) deficiency is an inherited rare bleeding disorder with an autosomal recessive manner. The prevalence of this disorder is about one in 2,000,000 people in general population, but it is more common in areas with a high rate of consanguinity. To date, there is no report on the absence of prothrombin, which is a life-threatening disorder.

Considering the importance of factor II in body homeostasis, this study aimed to find any possible mutation of coagulation factor II codons in patients with inherited factor II deficiency in southeastern Iran.

Materials and Methods: This study was conducted on 12 patients with inherited deficiency of prothrombin. Early diagnosis was based on clinical symptoms, laboratory evaluation, and family history. Then, the function level of prothrombin was measured, and the initial diagnosis of disease was confirmed, and polymerase chain reaction (PCR) analysis was performed. Finally, gene sequencing and genotyping of factor II was done.

Results: Molecular analysis indicated a point mutation in exon 7 in three patients and a frameshift mutation in exon 14 due to addition of a thymine base at position 1760-1761 in one patient, both of which have been reported for the first time.

Conclusions: Molecular methods performed on patients from Southeastern Iranian population in terms of coagulation factor II deficiency revealed a substitution mutation in exon 7 in three patients and a frameshift mutation in exon 14 in one patient, both of which were reported for the first time. Considering the significant difference between the clinical symptoms of the present study and previous studies, probably the type of mutations reported in this study (for the first time) caused these clinical symptoms, but statistical studies did not show any relationship between the type of mutation and the occurrence of clinical symptoms. And it needs more investigations on more patients, with a larger population.

Keywords: Factor II deficiency, Prothrombin, Congenital bleeding disorders, Blood coagulation factors

Introduction
Human prothrombin (factor II) is a 70 kDa glycoprotein synthesized in presence of Vitamin K in the liver. The gene encoding prothrombin (F2) is located on chromosome 1p11-q12 and contains 14 exons and 13 introns, including 20210 base pairs. Prothrombin molecule includes four domains, including Gla-domain, kringle-1, kringle-2, and canonical protease domain that contains A and B chains (1). After degradation of prothrombin complex by prothrombinase complex (consisting of factors X and V, calcium, and phospholipid), kringle-1 and 2 domains are released and active thrombin is formed. Thrombin includes two polypeptide chains A (36 amino acids) and B (259 amino acids). It is a bifunctional enzyme. On the one hand, it is involved in coagulation cascade via activation of a number of clotting factors (I, V, VIII, XI, XIII) and plays an anticoagulant role by activating thrombomodulin/protein C pathway (2,3).

Hereditary deficiency of this factor, which has a prevalence of approximately one in 2,000,000 people in general population, can manifest in mild or severe mode. This autosomal recessive disorder can affect all the human races. The cause of hereditary deficiency of factor II, either mild or severe, is often a form of gene mutation that involves the coding gene of this factor. So far, more than 50 mutations have been detected in factor II mainly in the form of missense mutation (80%). However, other mutations impair the structure of prothrombin, including insertion/deletion (10%), nonsense (6%), and splice site (4%) mutations.

Complete lack (absence) of prothrombin is contrary to human life and no report has been mentioned in this regard (4,5). Severe bleeding is the most obvious clinical manifestation in hypo-prothrombinemia with <10% plasma levels of prothrombin in homozygous patients. In studies on patients with prothrombin deficiency, intracerebral hemorrhage (ICH, which is life threatening), hemotoma, and spontaneous petechiae have been observed in 60% of patients, hemarthrosis in 42%, and gastrointestinal bleeding in only 12% of patients.
Bleeding after tooth extraction and menorrhagia in 20% of homozygous women with menorrhagia are common symptoms. In patients with this defect, routine coagulation tests such as PT (prothrombin time) and aPTT (activated partial thromboplastin time) are variably prolonged.

This disorder is approximately 10 times more common in regions with a high rate of consanguineous marriages (6,7). So far, no studies have been carried out on coagulation factor II in Iran. Nonetheless, some studies evaluated patients with thrombosis and their association with G20210A polymorphism in this factor. Due to high rates of consanguinity in Iran, the incidence of factor II deficiency, such as autosomal recessive inherited disease, is expected to be high. Considering the importance of this factor in body homeostasis, this study aimed to find any possible mutations of coagulation factor II codons in patients with inherited factor II deficiency in southeastern Iran.

Materials and methods

Study Design

In this study, 12 patients with factor II deficiency were selected from Ali Asghar Children's Hospital in Zahedan, Iran from 30 April to July 1, 2015. Routine coagulation tests of PT, PTT, and BT were performed for these patients. Prothrombin activity was measured in plasma for all subjects who had prolonged PT and PTT results; these subjects were candidates for determining the genotype of coagulation factor II codons.

Informed consent was obtained from all subjects. The questionnaires were completed by a physician through interviewing with the patients or their families (children under 16 years of age). The questionnaire included such information as age, gender, city of residence, age of diagnosis, family history, history of consanguineous marriage of parents, clinical symptoms, treatment, prophylaxis, and disease outcome. The study protocol was approved by the ethical committee of Tehran University of Medical Sciences, Iran.

The patients were divided into three groups based on the level of factor II activity as follows: <1%, 1-12%, and >12%. Moreover, based on clinical signs and bleeding symptoms, the patients were divided into four grades: Grade 0: patients without symptoms of bleeding, Grade I: patients showing clinical symptoms following injury or drug injection, Grade II: patients with minor spontaneous bleedings such as ecchymosis, bruising, bleeding gums, epistaxis, and bleeding from minor injuries, and Grade III: patients with severe life-threatening spontaneous bleeding, including cerebral bleeding, gastrointestinal bleeding.

Blood Sampling and Isolation of DNA

Whole blood samples with ethylenediaminetetraacetic acid (EDTA) anticoagulant were collected from the patients. Frozen blood samples were dispatched on dry ice to the Hematology Department of Tehran University of Medical Sciences to detect the mutations. EDTA containing plasma of patients was stored at -70°C until use. Genomic DNA of patients was extracted from EDTA containing peripheral blood samples using genomic DNA extract kit (Viogene Company, UK) and was kept at -20°C in aliquots.

PCR Analysis

The extracted DNAs were analyzed by PCR for genetic mutation in prothrombin gene. Oligonucleotide primers (Table 1) were chosen according to previous studies and synthesized by cDNA synthesis kit (Sinaclon, Iran). All the exons, introns, and 5'UTR and 3'UTR regions of factor II were amplified by PCR using Eppendorf thermocycler. PCR reaction was conducted using specific shuttle primers for each exon. Products of PCR were fractionated on 2% agarose gel electrophoresis and stained by ethidium bromide.

DNA Sequencing

Sequencing was based on the Sanger method, also known as chain termination. In this method, DNA is sequenced in a single pattern. After PCR, amplified DNA must be isolated from other agents. Glycogen, ammonium acetate, and absolute alcohol were used for this purpose, which caused DNA to separate and precipitate in the tube. All the steps of purification and sequencing were performed in Pishgam Company. After receiving the sequencing results, the sequences were analyzed with CodonCode Aligner software (version 5.1.5) and statistical data were performed using the Statistical Package for the Social Sciences (SPSS) software, version 18.

Prothrombin Activity Assay

The level of plasma activity of factor II in all patients included in the study was determined by Zahedan Hemophilia Center. Outcomes were expressed as a percentage of the quantity of prothrombin in pooled normal plasma, which was random designated as 100%.

Statistical Analysis

Chi-square test was used to evaluate the correlation between factor II and severity of bleeding symptoms as well as correlation between mutations and clinical symptoms. Data were analyzed using SPSS version 18. A P
value <0.05 was considered statistically significant.

**Results**

**Baseline Characteristics of Subjects**

In this study, out of 12 included patients, seven (55.83%) were male and five (41.66%) were female. The mean age of male and female patients was 13.4 (range: 3-21) and 55.2 (range: 47-65) years, respectively. The mean age of disease diagnosis was 4.5 and 44.6 years in males and females, respectively. According to the report by Hemophilia Center, plasma level of factor II was 7-14% in all the patients. None of the patients had <1% factor level, nine patients had 1-12% factor activity, and three patients had >12% factor activity.

In the present study, gynecological examination of female patients showed that all the studied patients had menorrhagia and postpartum hemorrhage (100%) and two of them had ICH hemorrhage (40%). Out of 12 patients, there were five cases of epistaxis (41.6%), two cases of bleeding after tooth extraction (16.6%), two cases of deep hematoma (16.6%), and five cases of ecchymosis (41.6%). Joint, gastrointestinal (GI), and ICH bleedings were not observed in any of the patients. The characteristics of patients under study is shown in Table 2. Figure 1 shows clinical symptoms of patients with hereditary factor II deficiency.

**Genetic Studies**

In three patients (No. 3, 5, and 7), a point mutation was found in exon 7. The guanine (G) base at position 844 was substituted by adenine (A). This homozygous mutation, which is reported for the first time, is known as p.282 asp>asn, indicating substitution of asparagine for aspartic acid in amino acid 282. Figure 2 shows this mutation.

In one patient (No. 11), frameshift mutation was observed in exon 14. In this mutation, due to addition of a thymine (T) base to exon between 1760 and 1761 positions, the stop codon appeared prematurely, causing

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**Table 1.** Primers Used for PCR Analysis of Factor II Codons

<table>
<thead>
<tr>
<th>Primer Type</th>
<th>Primer Sequence</th>
<th>Exon</th>
<th>Product Length</th>
</tr>
</thead>
<tbody>
<tr>
<td>Forward</td>
<td>CATGTTAGTCAACATTAACCCAGAGG</td>
<td>1</td>
<td>246</td>
</tr>
<tr>
<td>Reverse</td>
<td>GCCAGCTCTGTTGCTCTGTCT</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>GTTCCTAGGAGCTGCCTGTCCAFGAC</td>
<td>2</td>
<td>275</td>
</tr>
<tr>
<td>Reverse</td>
<td>CTCTGGAAAGTGCTGACGGTG</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>GTGAGGAGAGCAATAACCTTAAT</td>
<td>3</td>
<td>203</td>
</tr>
<tr>
<td>Reverse</td>
<td>CTGATGCTATGTCGACGAGCCAAG</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>CAACACATCCCCTACACCTAGTAC</td>
<td>4</td>
<td>171</td>
</tr>
<tr>
<td>Reverse</td>
<td>CTATGCTCCTGCTCCCCCAACCCC</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>GAAAATAGTCCCCAGGCTCCAAG</td>
<td>5</td>
<td>219</td>
</tr>
<tr>
<td>Reverse</td>
<td>GAACCTGAGGTACCTGCTCCTCCT</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>GCCGCTGGCGTGACCAACACAT</td>
<td>6</td>
<td>241</td>
</tr>
<tr>
<td>Reverse</td>
<td>AGGCCGGGGCCGTGCTGGGT</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>TGAGGAATGGCCGCTGCCAG</td>
<td>7</td>
<td>259</td>
</tr>
<tr>
<td>Reverse</td>
<td>CAGCAGTGCGAAGGCGAGCTTTG</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>AGCCAGGTGAGGTAGTGCGATCCCCG</td>
<td>8</td>
<td>249</td>
</tr>
<tr>
<td>Reverse</td>
<td>AGGGAGTCAGTGAGGGCGAGGTC</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>CTCACTCTCAGCTCCTTAATGCT</td>
<td>9</td>
<td>254</td>
</tr>
<tr>
<td>Reverse</td>
<td>AGGACTGAGACCCCTGCGACGAC</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>GTGAACCTCGACGTCTCCTATTCT</td>
<td>10</td>
<td>256</td>
</tr>
<tr>
<td>Reverse</td>
<td>TCTGCCCTCTAGCTAAACAGCAT</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>CAAGCCGGTGTCCTGTCGGGGTGGTG</td>
<td>11</td>
<td>254</td>
</tr>
<tr>
<td>Reverse</td>
<td>CTGGGCCCTACCTGTCCTCCT</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>TGGACTCTCACCAGCTGTGGTCCTGCT</td>
<td>12</td>
<td>281</td>
</tr>
<tr>
<td>Reverse</td>
<td>CAGCCAACCTGTTTCTAGATCATGAG</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Forward</td>
<td>CCTGAACTTGATCATCTATGGAAACC</td>
<td>13</td>
<td>242</td>
</tr>
<tr>
<td>Reverse</td>
<td>AGGGGATTCGGTGCTCAGGGCAGC</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

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**Figure 1.** Clinical Symptoms of Patients With Hereditary Factor II Deficiency.
a defective protein translation. It should be noted that this homozygous mutation has been observed for the first time. Figure 3 shows this mutation.

**Correlation Between Mutation And Clinical Features**

Clinical symptoms in patient No. 3 included menorrhagia, postpartum hemorrhage, and CNS bleeding. There was no correlation between mutation at position 844 of exon 7 and clinical symptoms ($P = 0.5$). The clinical symptoms in patient No. 5 included epistaxis and gum bleeding. There was no significant correlation between mutation in patient No. 5 and clinical symptoms ($P = 0.54$). Also, there was no significant correlation between mutation in position 844 of exon 7 and the level of factor II activity ($P = 0.24$), as well as severity of disease ($P = 0.5$).

Clinical symptoms in patient No. 11 were ecchymosis and nose bleeding. There was no significant correlation between frameshift mutation and nose bleeding, as well as ecchymosis ($P = 0.49$). Also, no significant correlation was found between frameshift mutation, severity of symptoms, and the level of factor II activity ($P = 0.55$).

Statistical studies did not show any relationship between mutation type and the incidence of clinical symptoms. Among clinical symptoms, a significant correlation was only found between factor II activity level, postpartum hemorrhage ($P = 0.018$), and menorrhagia ($P = 0.018$).

**Discussion**

This study was conducted on 12 patients with factor II deficiency from different cities in Iran in late 2016. It is believed that the actual quantity of patients with factor II deficiency is higher in this region because of the high rate of consanguineous marriages.

Congenital deficiency of factor II is a rare bleeding disorder with a prevalence of one in 2,000,000 people in general population. It is an autosomal recessive disorder affecting all the human races (8). This disorder is more prevalent (nearly 10 times higher frequency) in regions with a high propensity for consanguineous marriages. Caucasians, Middle East Asian immigrant population, as well as patients with Latinohispanic background (Barcelona, Padua, Segovia, Puerto Rico, etc) account for nearly 70% of patients with factor II deficiency (9).

Severe bleeding is the most obvious clinical manifestation in hypo-prothrombinemia with <10% plasma level of prothrombin in homozygous patients. In studies on patients with prothrombin deficiency, ICH, hematoma, and spontaneous petechiae have been reported in 60% of patients. ICH is extremely dangerous in these patients. Hemarthrosis has been reported in 42%
of patients while GI bleeding has been observed only in 12% of cases. The common symptoms also include bleeding after tooth extraction (in 36% of patients) and menorrhagia (in 20% of homozygous women). No symptoms are often observed in heterozygous cases of this disease with normal levels of prothrombin in plasma (40-60%) until tooth extraction or after surgery.

Congenital prothrombin deficiency was first described by Quick in 1947. Shapiro et al evaluated congenital dysprothrombinemia in a large family and reported that plasma prothrombin concentration and its biological activity was half the normal level; it was concluded that the disease is inherited in an autosomal recessive manner. In another study, O’Marcaigh et al investigated the functional features and analyzed Dharan (Arg271 His) and Corpus Christi (Arg 382 Cys) mutations, as well as hypo-prothrombinemia (10-13). In 2009, Lancellotti et al began their research on congenital defects of prothrombin. They concluded that approximately 49 mutations can cause defects in prothrombin in the form of hypo-prothrombinemia and dys-prothrombinemia, and that the lack of human prothrombin is incompatible with life (14). In 2013, Lancellotti et al again conducted studies on congenital prothrombin deficiency and found some new mutations on FII gene (15). In 2014, Bafunno et al detected a new mutation causing dys-prothrombinemia. The level of FII activity was 0.82% in this mutation, which occurred in the form of F2, c.1090 T/A (p.Val322Glu) and showed normal antigenicity. In this disorder, activation of mutated factor II by Okarin caused mesothrombin aggregation, which caused impaired prothrombin maturation towards thrombin. Minor bleeding was observed in this study (16).

Among 12 patients with factor II deficiency in this study, mutation was found in only four cases, of which three were point mutation and one case was frameshift mutation. Also, 33.33% of patients had considerable mutation. The findings of this study are different from previous studies in terms of clinical symptoms. In previous studies, ICH, hematomas, and spontaneous petechiae were the most common findings, while epistaxis and ecchymosis were the main clinical symptoms in the present study. Epistaxis has not been reported in previous studies. Bleeding from joints, GI tract or ICH were not reported in our patients. In previous studies, 20% of homozygous women showed signs of menorrhagia, whereas 100% of women in this study had menorrhagia. ICH was observed in two (40%) out of five women under study and postpartum bleeding was observed in 100% of women, while there was no report of this type of bleeding in factor II deficiency. Bleeding after tooth extraction was observed in 16.6% of patients in this study, but it was seen in 36% of patients in previous studies (17,18). Figure 4 compares the clinical manifestations of factor II deficiency in this study and previous studies.

Statistical analysis did not indicate any relationship between mutation type and clinical symptoms.

Limited number of patients, lack of cooperation from some patients, and high cost of sequencing are some of the limitations of the present study. It is recommended to do the following for detailed information: 1) repeat the study on a larger population of patients to find clinical symptoms and type of mutation, 2) evaluate the level of factor II at the level of mRNA and assess its relationship with the level of factor II antigenic, and 3) study the epigenetic
modification of factor II in terms of methylation and its relationship with expression level of factor II.

**Conclusion**

Molecular methods performed on patients from Southeastern Iranian population in terms of coagulation factor II deficiency revealed a substitution mutation in exon 7 in three patients and a frameshift mutation in exon 14 in one patient, both of which were reported for the first time. Considering the significant difference between the clinical symptoms of the present study and previous studies, probably the type of mutations reported in this study (for the first time) caused these clinical symptoms, but statistical studies did not show any relationship between the type of mutation and the occurrence of clinical symptoms. And it needs more investigations on more patients, with a larger population.

**Authors' Contribution**

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Methodology: Shaban Alizadeh, Hamed Soleimani Samarkhazan and Majid Naderi.

Validation: Hamed Soleimani Samarkhazan and Ziba Majidi.

Formal Analysis: Hamed Soleimani Samarkhazan and Zahra Kashani Khatib.

Investigation: Hamed Soleimani Samarkhazan and Shaban Alizadeh, Zahra Kashani Khatib and Ziba Majidi.

Resources: Hamed Soleimani Samarkhazan and Shaban Alizadeh.

Data Curation: Shaban Alizadeh and Majid Naderi.

Writing—Original Draft Preparation: Hamed Soleimani Samarkhazan and Ziba Majidi.

Writing—Review and Editing: Hamed Soleimani Samarkhazan and Ziba Majidi.

Visualization: Hamed Soleimani Samarkhazan, Zahra Kashani Khatib and Ziba Majidi.

Supervision: Shaban Alizadeh.

Project Administration: Shaban Alizadeh and Majid Naderi.

**Conflict of Interests**

The authors declare that there are no competing interests.

**Ethical Issues**

The study protocol was approved by the ethical committee of Tehran University of Medical Sciences, Iran.

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**References**